Ectopic expression of Pax4 in pancreatic δ cells results in β-like cell neogenesis

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The recent demonstration that pancreatic α cells can be continuously regenerated and converted into β-like cells upon ectopic expression of Pax4 opened new avenues of research in the endocrine cell differentiation and diabetes fields. To determine whether such plasticity was also shared by δ cells, we generated and characterized transgenic animals that express Pax4 specifically in somatostatin-expressing cells. We demonstrate that the ectopic expression of Pax4 in δ cells is sufficient to induce their conversion into functional β-like cells. Importantly, this conversion induces compensatory mechanisms involving the reactivation of endocrine developmental processes that result in dramatic β-like cell hyperplasia. Importantly, these β-like cells are functional and can partly reverse the consequences of chemically induced diabetes.

Introduction

The endocrine pancreas is organized into highly vascularized functional units, termed islets of Langerhans, encompassing five endocrine cell subtypes (α, β, δ, ε, and pancreatic polypeptide cells) responsible for the secretion of glucagon, insulin, somatostatin, ghrelin, and pancreatic polypeptide, respectively (Adrian et al., 1978; Roncoroni et al., 1983; Prado et al., 2004). Type 1 diabetes mellitus is a metabolic disease resulting from the autoimmune-mediated loss of insulin-producing β cells. Such loss induces a chronic hyperglycemia, which, left untreated, could have grave vascular consequences (Morrish et al., 2001; Alwan, 2010; Pascolini and Mariotti, 2012). Despite current therapies (mostly exogenous insulin supplementation), patients with type 1 diabetes mellitus exhibit an overall shortened life expectancy and an altered quality of life because of their inability to strictly regulate glucose homeostasis (World Health Organization, 2016). Therefore, in a search for alternative treatments, approaches aiming at inducing/gaining further insight into the molecular mechanisms underlying β cell (neo)genesis during pancreas morphogenesis and throughout adulthood are of growing interest. Consequently, several studies demonstrated that during pancreatic development, the cooperation of several transcription factors successively specifies endodermal progenitor cells toward the pancreatic, endocrine, and ultimately hormone-expressing cell fates. Among the latter, Arx and Pax4 mutually inhibit each other at the transcriptional level and thereby differentially specify the α and β/δ cell fates, respectively (Sosa-Pineda et al., 1997; Collombat et al., 2003). Interestingly, it was shown that the ectopic expression of Pax4, or the specific loss of Arx, in adult or embryonic α cells triggers their conversion into β-like cells and their subsequent neogenesis from the ductal epithelium (Collombat et al., 2009; Al-Hasani et al., 2013; Courtney et al., 2013). Importantly, such a cycle of neogenesis and conversion results in a high number of β-like cells that are functional and can revert the consequences of chemically induced diabetes multiple times. It is worth noting that in these different models, in addition to glucagon-expressing cells neogenesis, an increase in the number of somatostatin-expressing δ-like cells was consistently observed. Unexpectedly, although continuously detected, these δ cells were not found to accumulate over time. Based on these observations, and considering that δ cells are closely related to β cells during development, we sought to determine whether adult δ cells could not be induced to adopt a β-like cell identity. To address this question, we generated animals allowing the ectopic expression of Pax4 specifically in δ cells. Interestingly, our results provide evidence that adult δ cells can be reprogrammed into functional β-like cells upon the sole expression of Pax4. Importantly, we show a progressive islet neogenesis and islet hypertrophy together with dramatic endocrine cell hyperplasia. Together, our
analyses suggest that the δ-to-β-like cell conversion provokes a disruption of islet homeostasis triggering a reawakening of the endocrine differentiation program and resulting in a cycle of endocrine cell neogenesis and further δ-to-β-like cell reprogramming. Interestingly, upon chemically induced diabetes, transgenic mice ectopically expressing Pax4 in δ cells display an extended life span and a partial recovery of the β cell mass.

**Results**

**Generation and characterization of animals allowing the ectopic expression of Pax4 in somatostatin-expressing cells**

Aiming to determine whether the sole ectopic expression of Pax4 in δ cells could alter their phenotype/identity in vivo, we first crossed Sst-Cre animals (Fig. 1 A) with the ROSA26-β-gal mouse line (Soriano, 1999; Fig. 1 A). Our analyses of the pancreata from the resulting Sst-Cre::ROSA26-β-gal transgenic mice validated the specificity of Cre expression solely in somatostatin-producing cells (Fig. 1 C). Importantly, no glucagon-expressing cells were found positive for the β-galactosidase tracer, further confirming such δ cell specificity (Fig. 1 D). Subsequently, Sst-Cre animals were mated with Pax4-OE mice (Collombat et al., 2009; Fig. 1 B). In the resulting Sst-Cre::Pax4-OE double-transgenic mice, Pax4 ectopic expression was clearly detected in Cre-expressing somatostatin+ cells (Fig. 1, E–G). Accordingly, quantitative analyses confirmed such specificity with an ectopic expression of Pax4 in 66 ± 3.09% of somatostatin-expressing cells (Fig. 1, E and F). Importantly, Sst-Cre::Pax4-OE transgenic mice were found to be viable and fertile, and no premature death was observed. Along the same line, no statistical difference was observed in the glycemia of control and transgenic animals of matching ages (Fig. 1 H), demonstrating that the ectopic expression of Pax4 in somatostatin+ cells does not impact basal glycemia levels.

**Pax4 expression in δ cells results in progressive islet hypertrophy and insulin-producing cell hyperplasia**

The pancreata of Sst-Cre::Pax4-OE animals and age-/sex-matched controls were analyzed by immunofluorescence at 2, 5, and 8 mo of age. Importantly, an increase in islet number and size was evidenced in 2-mo-old Sst-Cre::Pax4-OE animals compared with controls (Fig. 2, A and B). As interesting were the further augmentations in both islet number and size observed in 5- and 8-mo-old Sst-Cre::Pax4-OE pancreata relative to controls (Fig. 2, C and D). These results were confirmed using quantitative analyses that demonstrated a significant increase in islet size (2.38 ± 0.43-fold) and insulin content (1.48 ± 0.27-fold) in 2-mo-old Sst-Cre::Pax4-OE pancreata compared with controls (Fig. 2, E and F). Important to note is the quantification of insulin+ cells per pancreas demonstrated that the insulin+ cell hyperplasia/multiplication mostly resulted from a significant increase in insulin+ cell hyperplasia with a 3.43 ± 0.09-fold increase in insulin+ cell numbers compared with controls (Fig. 2 G). Additional analyses performed in 5-mo-old animals ascertained a further increase reaching 3.30 ± 0.20-fold in the islet count, 2.27 ± 0.11-fold in islet size, and a massive 7.57 ± 0.08-fold in insulin+ cell counts as compared with controls (Fig. 2, E–G). Similar augmentations were also observed in 8-mo-old animals when compared with age-/sex-matched controls (Fig. 2, E–G), suggestive of a plateau in islet growth and multiplication after 5 mo of age. Focusing our quantitative studies on the relative endocrine cell proportions within hypertrophic islets versus controls, the insulin+ cell share was found slightly higher than normal in 5-mo-old Sst-Cre::Pax4-OE islets (Fig. 2 H), whereas the glucagon+ cell proportion was unchanged (Fig. 2 I). Surprisingly, the somatostatin+ cell share was 1.61 ± 0.65-fold reduced in 2-mo-old Sst-Cre::Pax4-OE islets and further diminished (3.05 ± 0.53-fold) in their 5-mo-old counterparts (Fig. 2 J). In addition to the evidently altered endocrine cell counts observed in Sst-Cre::Pax4-OE pancreata, an abnormal positioning of non-β cells was noted within the islets. Indeed, although glucagon- and somatostatin-expressing cells were predictably found uniformly distributed at the periphery of control islets (Fig. S1, A and B; and Video 1), a preferential localization at a pole of the islets, close to adjacent ducts, was evidenced for these non-β cells in 2-mo-old Sst-Cre::Pax4-OE islets (Fig. S1, C and D; and Video 1). Interestingly, these alterations appeared to result from a continued endocrine cell neogenesis processes. Accordingly, quantitative analyses of the total glucagon- and somatostatin-expressing cell counts per pancreatic sections revealed a progressive increase of both cell subpopulations in the pancreas of Sst-Cre::Pax4-OE transgenics (Fig. S1, E and F). Together, these results suggest that the ectopic expression of Pax4 in δ cells promotes a progressive increase in islet number, size, and cell content until 5 mo of age.

**Ectopic expression of Pax4 in somatostatin+ cells induces their conversion into β-like cells**

To investigate the fate/destiny of Pax4-expressing δ cells in Sst-Cre::Pax4-OE pancreata, further immunofluorescence experiments were undertaken. The expression of the β-galactosidase tracer was therefore monitored, comparing Sst-Cre::ROSA26-β-gal mice with Sst-Cre::Pax4-OE animals. In Sst-Cre::ROSA26-β-gal control pancreata, β-gal+ cells were found, as expected, in the mantle of the islets, where somatostatin-expressing cells are normally located (Fig. 3 A). Although no insulin/β-gal double-positive cells could be found in control islets (Fig. 3 A), several insulin+/β-galactosidase+ cells were detected in 2-mo-old Sst-Cre::Pax4-OE pancreata (Figs. 3 B and S2 A). Interestingly, similar lineage tracing experiments performed on 5-mo-old transgenics revealed a further increased number of insulin+ cells labeled with β-galactosidase (Figs. 3 C and S2 B). These results were confirmed using quantitative analyses. Indeed, although no insulin+ cells expressed β-galactosidase in control islets, a proportion of 3.1 ± 0.36% of insulin+ cells expressed β-galactosidase in 2-mo-old transgenic islets (Fig. 3 D). Similarly, 5.1 ± 0.69% of insulin+ cells were labeled with β-galactosidase in 5-mo-old transgenic islets (Fig. 3 D). To investigate the mechanisms by which Pax4 could induce such conversion, we sought to assess the expression of several δ cell markers in mice with Sst-Cre::Pax4-OE transgenics (Zhang et al., 2014). Together, findings demonstrate that Pax4 expression in somatostatin+ cells induces their conversion
Pax4 induces δ-to-β-like cell conversion

Druelle et al. 4301

into β-like cells, presumably trough Hhex inhibition. Unfortunately, further in-depth analyses are impossible because of the lack of a proper δ cell line (for in vitro studies) and the small number of such cells (for in vivo assessment). Importantly, and considering that the insulin+ cell count per islet massively increases with age, our results suggest that the proportion of insulin+ cells with a δ cell ontogeny increases proportionally with the islet growth. As important is the observation that most neo-generated insulin+ cells are not derived from δ cells, indicating that additional processes are at play.

**Ectopic expression of Pax4 in δ cells results in increased proliferation within the ductal epithelium.**

Our results demonstrate that the ectopic expression of the Pax4 gene in δ cells is sufficient to induce their reprogramming into cells displaying an insulin+ cell identity. However, this conversion can clearly not account for the massive endocrine cell hyperplasia observed in Sst-Cre::Pax4-OE animals, suggestive of the involvement of additional mechanisms. Thus, to investigate whether the latter could imply an increase in cell replication, cell proliferation rates were assayed in Sst-Cre::Pax4-OE mice and matched controls treated with either iodo-deoxyuridine (IdU) or BrdU for different durations. Importantly, after a short IdU pulse (3 d), a significant augmentation in replicating cell numbers was observed in the pancreas of 2-mo-old transgenics, as indicated by increased IdU+ cell numbers (Fig. 4, A and E). Strikingly, most proliferating cells were located not within the islets of Langerhans but rather within the “ductal lining” (referring to cells located in the ductal epithelium and its immediate vicinity; Fig. 4 E). Aiming to monitor their outcome, pulse-chase experiments (3 d of IdU labeling followed by 7 d of chase) were performed as a means of short-term lineage tracing, as previously reported (Alcolea and Jones, 2014; Fink et al., 2015; Hsu, 2015). Importantly, after a 7-d chase, these IdU-labeled cells were found not only in the ductal lining of 2-mo-
old Sst-Cre::Pax4-OE pancreata but also within the islets of Langerhans (Fig. 4, B and F). Accordingly, longer IdU treatment (10 d) revealed numerous IdU+ cells in the ductal epithelium and lining (Fig. 4, C, D, and G; and Fig. S3, A–F), and IdU+ cells were also found within the islets of Sst-Cre::Pax4-OE mice (Fig. 4, C D, and H), further suggesting that proliferating cells migrated from the ductal compartment to the islets of Langerhans. Of note, the slow rate of proliferation in control ducts was also observed in larger ducts (Fig. S3, A–C), whereas increased ductal proliferation was also detected in smaller ducts in 2-mo-old Sst-Cre::Pax4-OE transgenics (Fig. S3, D–F), indicating that the increase in the proliferation of duct-lining cells in transgenic pancreata does not seem to depend on the size of the ducts. Quantitative analyses verified these observations and outlined a 5.73 ± 0.38-fold increase in the numbers of IdU+ cells solely within the ductal lining or epithelium after short-term IdU treatment comparing Sst-Cre::Pax4-OE pancreata and controls (Fig. 4 I). Chasing the label for 7 d resulted in a 3.93 ± 0.41-fold augmentation in IdU+ cell counts within the endocrine compartment of 2- and 5-mo-old mutant islets, whereas a 4.10 ± 1.04-fold increase was still detected within the ductal lining/epithelium. Similar results were obtained after 10 d of IdU treatment with a 2.47 ± 0.14-fold increase in the numbers of IdU+ cells within the endocrine compartment (Fig. 4 I). Of note, no difference in IdU+ cell counts was observed in the acinar tissue comparing both groups (Fig. 4 I). Concomitant with the plateau in islet hypertrophy observed after 5 mo of age, comparative quantifications of endocrine and nonendocrine cell proliferation rates in 5- and 8-mo-old mutants did not reveal any differences compared with controls (n = 7; I). A significant decrease in somatostatin+ proportions was observed in 2- and 5-mo-old mutant islets compared with controls (n = 7; I). All values are depicted as mean ± SEM. Statistics were performed with the Mann–Whitney test or unpaired t test with Welch’s correction. ***, P < 0.001; **, P < 0.01; *, P < 0.05. Bars, 500 µm. INS/Ins, insulin.

Reexpression of Neurog3 and epithelial-to-mesenchymal transition (EMT) markers in Sst-Cre::Pax4-OE pancreata

Aiming to gain further insight into the mechanisms underlying ductal cell proliferation and the resulting endocrine cell neogenesis observed in Sst-Cre::Pax4-OE pancreata, we wondered whether these processes could involve a reawakening of the developmental endocrine differentiation program that would
result in the neogenesis of all endocrine cell subtypes as seen during embryogenesis. Thus, to test this hypothesis, the expression of the proendocrine/developmental master gene Neurog3 was investigated. Whereas Neurog3 was expectedly not found expressed in adult control pancreas (Fig. 5 A), a reexpression of this proendocrine gene was evidenced in several ductal cells and within the ductal lining (Fig. 5 B). Because of known difficulties to detect Neurog3 expression by immunofluorescence and to ascertain the reexpression of this proendocrine gene, total pancreas quantitative RT-PCR analyses were performed. A 2.26 ± 0.30-fold increase in Neurog3 transcripts was demonstrated in 2-mo-old Sst-Cre::Pax4-OE pancreata, further confirming its reexpression (Fig. 5 K). Similarly, a 3.40 ± 1.19-fold increase in the expression of the Neurog3 target IA1 was observed (Fig. 5 K). Combined, these observations support the notion of a reawakening of the expression of Neurog3 in the ductal lining of adult animals expressing Pax4 in δ cells. Importantly, Neurog3+ cells give rise to all endocrine cells during development, further suggesting that the islet hypertrophy observed Sst-Cre::Pax4-OE could be caused by a reenactment of the developmental endocrine differentiation program. Aiming to further ascertain this hypothesis, we focused on EMT, a key developmental process. EMT is orchestrated by a cascade of events, including the loss of an epithelial cell identity and the acquisition of a mesenchymal cell phenotype and associated markers (such as vimentin, nestin, Sox11, Snail2, and N-cadherin; Chiang and Melton, 2003). We therefore assessed the expression of an EMT hallmark, Vimentin, in 2-mo-old Sst-Cre::Pax4-OE pancreata and controls. Surprisingly, our analyses revealed a massive increase in the numbers of vimentin+ cells in the ductal epithelium, ductal lining, and close to adjacent islets (Fig. 5, C–J). Total pancreas quantitative RT-PCR analyses further confirmed these observations when focusing on additional EMT markers, with a 1.8-fold increase in Snail2 transcripts, a 15-fold increase in Nestin expression, and an eightfold augmentation of Sox11 transcripts in 2-mo-old transgenic pancreata compared with controls (Fig. 5 K). To verify that no inflammation or fibrosis occurred in the pancreas of transgenic mice, hematoxylin and eosin and Sirius red colorations obtained did not show any abnormalities, collagen deposits, or inflammatory infiltration (Fig. S4, A–D). In addition, to verify whether vimentin+ cells could correspond to pancreatic stellate cells, we focused on analyses on activated stellate cell markers, such as α-SMA or ICAM-1. None of these marker genes were detected in Sst-Cre::Pax4-OE pancreata, thus excluding a putative activation of pancreatic stellate cells (Fig. S4, E–H). Interestingly, further examination of Sst-Cre::Pax4-OE animals treated for 10 d with BrdU also unraveled several duct-lining BrdU+ cells expressing Vimentin (Fig. 5, E and F), suggesting that proliferating ductal cells undergo EMT. Additional analyses of 2-mo-old transgensics and controls revealed the presence of vimentin/synaptophysin+ cells located close to ducts (Fig. 5, G and H), indicating that cells undergoing EMT adopt an endocrine cell identity. Accordingly, vimentin+/β-galactosidase+ cells were also detected within the ductal lining of 2-mo-old.
Sst-Cre::Pax4-OE transgenics (Fig. 5, I and J). Together, these results support the notion that Pax4 ectopic expression in δ cells eventually induces a reawakening of endocrine developmental processes with the reexpression of Neurog3 in duct-lining cells and associated developmental processes, such cells eventually adopting an endocrine cell identity. Indeed, although ductal cell lineage tracing is impossible in the present context, the data accumulated suggest that proliferating ductal cells eventually adopt an endocrine cell identity by reenactment of the endocrine developmental program.

**Neo-generated insulin-expressing cells in Sst-Cre::Pax4-OE pancreata display a β-like cell phenotype and are functional.**

To further investigate the identity of the neo-generated insulin-expressing cells observed in Sst-Cre::Pax4-OE pancreata, thorough marker gene analyses were performed in 2-, 5- and 8-mo-old animals. In all three instances, our data indicated that all insulin+ cells (preexisting and neo-generated) displayed a β-like cell phenotype. Indeed, these cells uniformly expressed the bona fide β cell labels, such as PC1/3 (Fig. 6, A–D), Glut-2 (Fig. 6, E–H), Nkx6.1 (Fig. 6, I–L), Pdx1 (Fig. 6, M–P), and the pan-endocrine genes NeuroD1 (Fig. 6, Q–T) and Pax6 (Fig. 6, U–X). To assess the function of Sst-Cre::Pax4-OE insulin+ cells, intraperitoneal glucose tolerance tests were performed on 2-mo old Sst-Cre::Pax4-OE mice and matched controls. Interestingly, transgenic mice displayed an improved response with a lower peak in glycemia and a faster return to normoglycemia than controls (Fig. 7 A), suggestive of an increased functional β-like cell mass. In addition, insulin tolerance tests did not reveal any significant difference between 2-mo-old Sst-Cre::Pax4-OE animals and matched controls (Fig. 7 B), indicating that despite an increased β-like cell content, Sst-Cre::Pax4-OE mice do not develop any insulin resistance. These analyses led us to conclude that the hyperplastic insulin-expressing cells observed after the ectopic...
Pax4 induces δ-to-ß–like cell conversion

• Druelle et al. 4305

expression of Pax4 in somatostatin+ cells exhibit a β-like cell phenotype and are functional.

β-like cell neogenesis in Sst-Cre::Pax4-OE animals subjected to β cell ablation

The neo-generated β-like cells observed in Sst-Cre::Pax4-OE pancreata were further investigated in the context of chemically induced β cell ablation (Fig. 8 A). A single dose of 115 mg/kg streptozotocin (STZ) was administered to 2.5-mo-old transgenics and matching controls to induce the destruction of a vast majority of β cells without complete ablation to avoid predictable death (Fig. 8 B and E). As expected, after specific β cell loss, control and Sst-Cre::Pax4-OE mice developed hyperglycemia within 1 wk after injection (Fig. 8 H). Three months after injection, all control mice showed a glycemia >600 mg/dl and a high rate of lethality (41%), whereas transgenic mice displayed an average glycemia of 350 mg/dl (Fig. 8 H), suggesting a partial β cell mass restoration and an extended life span. This was further supported by the detection of insulin-expressing cells in islets of control and transgenic mice at different times after STZ-mediated β cell ablation. Indeed, although after 1 wk of STZ administration, control and Sst-Cre::Pax4-OE animals displayed a massive loss of insulin-expressing cells (Fig. 8, B and E), a progressive restoration of such cells was observed in transgenic mice (Fig. 8, F and G), and this partial recovery was absent in controls (Fig. 8, C and D). To further assess these regeneration processes, the β cell mass was quantified in control and transgenic pancreata 3 mo after STZ administration. Although decreased compared with their noninjected counterparts, STZ-treated Sst-Cre::Pax4-OE islets exhibited 16 ± 34% insulin-expressing cells compared with 8 ± 17% in treated-control islets (Fig. 8 I). Lineage tracing experiments were performed on STZ-injected control and transgenics, and we thereby detected insulin+/β-galactosidase+ cells in Sst-Cre::Cl−Pax4-OE pancreata (Fig. 8, J and K). To get more insight into the mechanisms involved in this partial β cell mass restoration, we assessed the expression of the mesenchymal marker vimentin. Interestingly, an important augmentation in the number of vimentin-expressing cells was observed in Sst-Cre::Pax4-OE transgenics 1 mo after STZ administration when compared with matched-controls (Fig. S5, A–F). In addition, we detected synaptophysin+ cells close to the mesenchymal marker expression.
ductal epithelium of STZ-injected Sst-Cre::Pax4-OE, and some of these cells expressed vimentin (Fig. S5, C and D). Of note, the assessment of cell proliferation after β cell ablation by the detection of Ki-67 protein did not reveal any difference in the number of replicating cells in the endocrine compartment when comparing controls and transgenic mice (Fig. S5, E and F), thereby excluding an increase in islet cell proliferation. Together, these results demonstrate that δ cells can be converted into functional β-like cells, and such conversion results in compensatory developmental endocrine cell neogenesis. Importantly, the β-like cells thereby generated can partly reverse the consequences of chemically induced diabetes, resulting in an extended life span.

**Discussion**

Here, we report that the ectopic expression of Pax4 in δ cells induces their conversion into cells displaying a β cell phenotype, and this induces compensatory mechanisms involving the mobilization of putative duct-lining cells that reenact developmental processes before differentiation into endocrine cells. Importantly, this cycle of neogenesis and further δ-to-β-like cell reprogramming results in a massive and progressive increase in the number of insulin-producing cells, resulting in islet hypertrophy and improved glucose tolerance. Equally important was the finding that neo-generated β-like cells are functional
and able to partly replace their endogenous counterparts upon chemically induced diabetes.

The ectopic expression of Pax4 in δ cells induces their conversion into β-like cells

Our results demonstrate that the ectopic expression of Pax4 in δ cells is sufficient to induce their efficient conversion into β-like cells. Indeed, we show that 3–5% of insulin+ cells arise from δ cells in 2- and 5-mo-old Sst-Cre::Pax4-OE animals (displaying a progressing insulin+ cell hyperplasia), suggesting a continuous processes of δ cell neogenesis and conversion into insulin+ cells. According to a previous study, near-total β cell ablation in mice results in the conversion of pancreatic δ cells into insulin-producing cells, and this conversion occurs only in juveniles (before 2 mo of age; Chera et al., 2014). Here, we demonstrate that upon Pax4 expression, somatostatin-expressing cells retain the ability to be converted into β-like cells until at least 5 mo of age. Whether this arrest is caused by a loss in δ cell plasticity or a negative feedback loop controlling the total endocrine cell content would require the establishment of an inducible system. In addition, we provide evidence that the conversion of δ cells into β-like cells can trigger a massive endocrine cell neogenesis, with most of these newly formed cells not displaying a δ cell ontogeny. Furthermore, although the presence of somatostatin+/Hhex+ cells within transgenic islets provides exciting insights into the mechanism by which Pax4 induces the conversion of somatostatin-expressing cells, it is impossible to know whether Hhex expression is down-regulated as a consequence of Pax4 ectopic expression or the δ-to-β-like cell conversion. It is indeed important to keep in mind that δ cells are continuously regenerated and turned into β-like cells in Sst-Cre::Pax4-OE mice that are less than 5 mo of age. One could bypass these concerns using a δ cell line in which Pax4 expression is forced. However, to the best of our knowledge, there is no δ cell line solely expressing the somatostatin hormone, not counting the fact that cell lines rarely retain their in vivo plasticity. The generation and analysis of gain-of-function/loss-of-function mutant animals for Hhex or the generation and analysis of animals expressing Pax4 in a conditional and inducible fashion in δ cells represent interesting alternatives.

Recruitment of Neurog3-reexpressing ductal cells and EMT reawakening

In this study, we demonstrate that most neo-generated β-like cells are not derived from δ cells. In fact, our analyses indicate that the δ-to-β-like cell conversion triggers compensatory mechanisms involving an increased proliferation of duct-lining cells, and such cells eventually adopt an endocrine cell identity. Accordingly, we outline a reexpression of the developmental gene Neurog3 within or close to the ductal lining upon the ectopic expression of Pax4 in somatostatin-expressing cells. Although lineage tracing of ductal cells would undeniably demonstrate the ductal ontogeny of the endocrine cell neogenesis observed in Sst-Cre::Pax4-OE animals, one may easily comprehend that it is impossible in the present context, as these transgenic animals already harbor a Cre recombinase–containing transgene. Indeed, to the best of our knowledge, the only available or working mouse lines allowing the lineage tracing of Ngn3- or vimentin-expressing cells rely on the Cre-Lox approach. Because of this technical limitation, we therefore resorted to short-term tracing of proliferating ductal cells combined to immunofluorescence detection of Ngn3 (and EMT makers) and quantitative PCR assessment of Ngn3 (and EMT makers). It is worth noting that this approach has been successfully used by others (Alcolea and Jones, 2014; Fink et al., 2015; Hsu, 2015), and adult Neurog3-reexpressing cells have been repeatedly described as endocrine progenitors involved in β cell regeneration (Xu et al., 2008; Al-Hasani et al., 2013; Pan et al., 2013). In addition,
and emphasizing that Neurog3 is responsible for the specification of all the endocrine cell lineages following specific ratios, the generation of endocrine cell types from the adult ductal epithelium would be expected to respect the standard endocrine proportions (≈80% of β cells/≈10% of α cells/≈10% of δ cells). Moreover, it has been shown that during mouse pancreas morphogenesis, Neurog3 induces the delamination of progenitor cells from the ductal epithelium through EMT processes (Gouzi et al., 2011). Interestingly, our data outline a massive increase in the expression of mesenchymal markers in the ductal lining of Sst-Cre::Pax4-OE transgenics, further supporting the idea that endocrine cell neogenesis putatively originates from the ductal epithelium. Altogether, our results clearly suggest that Pax4 expression in δ cells induces their conversion into β-like cells, subsequently activating a cycle of endocrine cell neogenesis, with the neo-generated δ cells being yet again converted into β-like cells (Fig. 9).

**Partial restoration of the β cell mass upon β cell ablation**

The ectopic expression of Pax4 in δ cells induces β-like cell neogenesis after chemically induced β cell ablation, resulting in a partial β cell mass restoration. Although not exhibiting euglycemia, Sst-Cre::Pax4-OE transgenics displayed a maximal glycemia of ≈350 mg/dl and an extended life span compared with controls. The reason underlying this incomplete recovery remains unclear but could potentially involve a slow regeneration processes or a loss of δ cell plasticity with ageing. Supporting the latter, the analyses of Sst-Cre::Pax4-OE animals suggest that δ cells eventually lose their plasticity at ≈5 mo of age. Obviously, it would be extremely interesting to determine whether such loss of regeneration potential is truly age dependent or the result of the continued endocrine neogenesis observed in Sst-Cre::Pax4-OE transgenics. The answer to this question will unfortunately have to wait for the generation of transgenic mice allowing the inducible expression of the Cre gene.
**Materials and methods**

**Ethics statement**

Animal protocols were reviewed and approved by an institutional ethics committee (Ciepal-Azur) at the University of Nice, and all colonies were maintained following European animal research guidelines. This project received approval from our local ethics committee (NCE/2011-22).

**Animal procedures**

Experiments were performed on transgenic and control males only. Controls included wild-type mice or transgene-negative litters. To assess cellular proliferation, animals were treated with BrdU or IdU in the drinking water for 3 or 10 d before examination (1 mg/ml solution). All mice were housed on a standard 12:12-h light/dark cycle, with standard diet food and water ad libitum.

**Generation of the Sst::Cre-Pax4-OE transgenic line**

Using the Cre-LoxP system to investigate δ cell plasticity in pancreatic islets, Sst-Cre::Pax4-OE animals were obtained by crossing the Sst-Cre mouse line (harboring a transgene encompassing the somatostatin promoter driving the expression of the phage P1 Cre recombinase; Taniguchi et al., 2011) with the previously described Pax4-OE line (whose transgene encompasses the ubiquitous CAG promoter upstream of a “GFP-STOP” cassette flanked by LoxP sites and followed by the Pax4 cDNA in front of an IRES and the β-galactosidase reporter (Collombat et al., 2009). In the resulting bitransgenic line, Pax4 is ectopically overexpressed in somatostatin-expressing cells and lineage of these cells can be traced.

**Histology and immunofluorescence**

Tissues were fixed in 4% paraformaldehyde for 30 min at 4°C, embedded in paraffin and 8 µm sections applied to glass slides. These sections were assayed as described previously (Collombat et al., 2003) using DAPI as counterstain. The primary antibodies used were guinea pig anti-insulin (1/500; DAKO), anti-Pdx1 (1/1,000; provided by C. Wright, Vanderbilt University, Nashville, TN); chicken anti-vimentin (1/4,000; Millipore); rat anti-somatostatin (1/250; Millipore) and anti-BrdU (1/50; Abcam); goat anti-somatostatin (1/500; Santa Cruz); mouse anti-BrdU (1/50; Roche), anti-Ngn3 (1/250; DSHB), and anti-glucagon (1/500; Sigma); rabbit anti-glucagon (1/500; R&D Systems), anti-glut-2 (1/500; Chemicon), anti-PC1/3 (1/500; Millipore), anti-Nkx6.1 (1/3000; NovoNordisk), anti-Pdx6 (1/500; BioLegend), anti-Pax4 (1/500; provided by B. Sosa-Pineda, Northwestern University, Evanston, IL), anti-NeuroD1 (1/500; Millipore), and anti-synaptophysin (1/400; Abcam). The secondary antibodies (1/1,000, Invitrogen and

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*Figure 9.* Schematic illustrating the consequences of Pax4 ectopic expression in somatostatin-expressing δ cells. Upon Pax4 expression in δ cells (1), these cells are converted into β-like cells (2 and 3). This leads to a disruption of islet homeostasis, mainly because of a local somatostatin shortage, further inducing the mobilization of ductal precursor cells [4], with such precursors reexpressing the proendocrine gene Neurog3 [5]. The latter cells undergo EMT before giving rise to insulin+, glucagon+, and somatostatin+ cells in normal/developmental proportions [6], and neogenerated somatostatin+ cells are yet again converted into β-like cells. Such a continuous cycle of regeneration and conversion results in insulin+ cell hyperplasia, islet hypertrophy, and islet neogenesis [7].
for normalization purposes.

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JCB • Volume 216 • Number 12 • 2017

manufacturer’s instructions. PCR reactions and detection were performed on a Mastercypher ep reaplex cycler using GAPDH as an internal control for normalization purposes.

β-Galactosidase–based lineage-tracing experiments

Pancreatic tissues were isolated and fixed for 30 min at 4°C in a solution containing 1% formaldehyde, 0.2% glutaeraldehyde, and 0.02% NP-40 and washed three times in cold PBS. The tissues were dehydrated in PBS-25% sucrose overnight at 4°C. Tissues were washed 2 h in Jung freezing medium (Leica Biosystems) and embedded in freezing medium on dry ice. 12- to 16-μm sections were applied to glass slides, dried, and washed three times in PBS. After 45-min blocking in PBS-FCS 5%, sections were incubated with a rabbit anti-β-galactosidase primary antibody (1/8,000; MP-Cappel).

Microscope image acquisition

For immunofluorescence analyses, images were collected at room temperature on an AxioImager.Z1 upright microscope (Zeiss) equipped with a Zeiss 10x Plan-Neofluar dry NA 0.3 and/or Zeiss 20× Plan-Apochromat dry NA 0.8 and/or Zeiss 20× Plan-Neofluar dry NA 0.5, a DAPI filter (excitation [Ex] 365/12 nm, dichroic mirror [DM] 395 nm, emission [Em] 445/50 nm), a GFP filter (Ex 470/40 nm, DM 495 nm, and Em 525/50 nm), a rhodamine filter (Ex 546/12 nm, DM 560 nm, and Em 608/65 nm), a Texas red filter (Ex 560/40 nm, DM 582 nm, and Em 630/785 nm), and/or a Cy5 filter (Ex 640/30 nm, DM 660 nm, Em 690/50 nm). For mosaic acquisitions, the microscope was equipped with an automated xy stage (Marzhauser). Images were acquired with a monochrome MRm axiocam camera (Zeiss) controlled with Axiosvision 4.9.1 software (Zeiss). Posttreatment image acquisition and analysis were performed using a homemade macro on Fiji (Schindelin et al., 2012). For histological analyses, images were collected at room temperature on an Axioplan2 upright microscope (Zeiss) equipped with a Zeiss 10x Plan-Neofluar PH1 dry NA 0.3 objective lens. Images were acquired with a color Infinity3-6 camera (Lumenera Corporation) controlled with Micro-Manager 1.48 software (Edelstein et al., 2014).

Tolerance tests and blood glucose level measurements

For challenge purposes, animals were fasted for 16–18 h and injected intraperitoneally with 2 g/kg body weight glucose or 0.75 U/kg insulin. Blood glucose levels were measured at the indicated time points postinjection with a ONETOUGH Vita glucometer (Life Scan, Inc.).

Induction of STZ-mediated diabetes

To induce hyperglycemia, STZ (Sigma) was dissolved in 0.1 M sodium citrate buffer, pH 4.5, and a single 115-mg/kg dose was administered intraperitoneally within 10 min of dissolution. Diabetes progression was assessed by monitoring the blood glucose levels and/or survival rates of mice.

RNA analyses

Mice were euthanized by cervical dislocation and the pancreatic tissue was isolated and placed in cold RNA later solution (Thermo Fisher Scientific) for 48 h at 4°C. RNA was isolated (RNAeasy; QIAGEN), and cDNA synthesis (SuperScript choice system; Invitrogen) was performed according to the manufacturer’s instructions. Quantitative RT–PCR was performed using validated primers (QIAGEN) and the QuantiTect SYBR Green RT-PCR kit (QIAGEN) following the manufacturer’s instructions. PCR reactions and detection were performed on a Mastercypher ep reaplex cycler using GAPDH as an internal control for normalization purposes.

Quantifications analyses

The entire pancreata of at least three mice per genotype and per condition were serially cut in 8-μm-thick sections and applied to glass slides, and every 5–10 pancreatic sections were processed for immunofluorescence. For Cre recombinase efficiency and β-galactosidase–expressing cells, positive cells were counted manually. IDU and BrdU counting was assessed manually by counting proliferative cells in the different pancreatic compartments. Images were processed as described in Microscope image acquisition. For hormone quantifications, widefield multichannel tile scan image treatment and analysis were done using Fiji (Schindelin et al., 2012) and a homemade semiautomated macro. In brief, the overall pancreas slice area was determined and measured using a low-intensity–based threshold on hormone labeling channel. Islets were then segmented and counted on the insulin labeling, applying a high-intensity threshold. In a second step, the respective area of each segmented islet was measured. The corresponding contours were then transferred to the somatostatin or glucagon images. Finally, the signal coverage of either glucagon or somatostatin was measured and then normalized to the total area of the corresponding islet.

Data analysis

All values are depicted as mean ± SEM of data from at least three animals. Data were analyzed using GraphPrism 6 software. Normality was tested using a D’Agostino–Pearson omnibus normality test, and appropriate statistical tests were performed. Results are considered significant if P < 0.0001 (***, P < 0.001 (**), P < 0.01 (*), and P < 0.05 (*)

Online supplemental material

Fig. S1 shows that the ectopic expression of Pax4 in δ cells results in endocrine cell neogenesis and an abnormal positioning of non-β cells within the islets of Sst-Cre::Pax4-OE transgenic mice. Data were analyzed using GraphPrism 6 software. Normality was tested using a D’Agostino–Pearson omnibus normality test, and appropriate statistical tests were performed. Results are considered significant if P < 0.0001 (***, P < 0.001 (**), P < 0.01 (*), and P < 0.05 (*).
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