The Phosphatase Dusp7 Drives Meiotic Resumption and Chromosome Alignment in Mouse Oocytes

Graphical Abstract

Highlights

- The phosphatase DUSP7 is an essential regulator of meiotic resumption
- DUSP7 dephosphorylates and inactivates cPKC isoforms
- Inactivation of cPKC isoforms is essential for the timely activation of Cdk1/Cyclin B

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In Brief

Tischer and Schuh study how oocytes resume meiosis after a prolonged prophase arrest. They show that the phosphatase DUSP7 drives meiotic resumption by dephosphorylating and inactivating conventional protein kinase C (cPKC) isoforms. Hyperactivation of cPKC isoforms in the absence of DUSP7 causes a decrease in the activity of the cell-cycle-regulating kinase Cdk1/Cyclin B, preventing meiotic resumption.
The Phosphatase Dusp7 Drives Meiotic Resumption and Chromosome Alignment in Mouse Oocytes

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SUMMARY
Mammalian oocytes are stored in the ovary, where they are arrested in prophase for prolonged periods. The mechanisms that abrogate the prophase arrest in mammalian oocytes and reinitiate meiosis are not well understood. Here, we identify and characterize an essential pathway for the resumption of meiosis that relies on the protein phosphatase DUSP7. DUSP7-depleted oocytes either fail to resume meiosis or resume meiosis with a significant delay. In the absence of DUSP7, Cdk1/CycB activity drops below the critical level required to reinitiate meiosis, precluding or delaying nuclear envelope breakdown. Our data suggest that DUSP7 drives meiotic resumption by dephosphorylating and thereby inactivating cPKC isoforms. In addition to controlling meiotic resumption, DUSP7 has a second function in chromosome segregation: DUSP7-depleted oocytes that enter meiosis show severe chromosome alignment defects and progress into anaphase prematurely. Altogether, these findings establish the phosphatase DUSP7 as an essential regulator of multiple steps in oocyte meiosis.

INTRODUCTION
Mammalian oocytes develop over a long period, spanning up to several decades in humans. Their development starts before birth in the embryo. The oocytes are initially very small and are stored for long periods in the ovary. Periodically, some oocytes become activated. They grow in size and accumulate mRNAs stored for long periods in the ovary. Periodically, some oocytes are expressed (Levy-Nissenbaum et al., 2003), but its physiological role is still unknown. When the oocytes have reached their full size, they are capable of maturing into a fertilizable egg. This maturation is triggered once every menstrual cycle by a surge of luteinizing hormone (LH). The surge of LH leads to the activation of the important cell-cycle regulator Cyclin-dependent kinase 1 (Cdk1/CycB). This complex is also known as the mitotic spindle assembly factor (Camps et al., 1998). Little is known about the function of DUSP7. In acute leukemia, DUSP7 is frequently over-expressed (Levy-Nissenbaum et al., 2003), but its physiological role is still unknown.

In mammalian meiosis (Pfender et al., 2015). Through this screen, we identified dual-specificity phosphatase (DUSP) 7 as a phosphatase that is essential for the resumption of meiosis.

Failure to exit the meiotic arrest has been implicated in female infertility (Hsieh et al., 2007; Mitri et al., 2014; Richards et al., 2002). Understanding the mechanisms that control meiotic resumption is thus of medical relevance. Current research on meiotic resumption mainly focuses on protein kinases and the protein phosphatases P1 and P2A (Adhikari and Liu, 2014; Holt et al., 2013). We recently performed a large-scale RNAi screen to identify genes that are involved in the regulation of mammalian meiosis (Pfender et al., 2015). Through this screen, we identified dual-specificity phosphatase (DUSP) 7 as a phosphatase that is essential for the resumption of meiosis.

DUSP7 is a cytoplasmic phosphatase and was identified as mitogen-activated protein kinase-phosphatase (MKP). DUSP7 is known to dephosphorylate the p42/p44 mitogen-activated protein kinases (p42/p44 MAPKs, better known as ERK1/2), which inactivates them (Caunt et al., 2008). Generally, DUSPs bind to their targets via a conserved arginine-rich kinase interaction motif, which is located amino-terminally of the phosphatase domain (Camps et al., 1998). Little is known about the function of DUSP7. In acute leukemia, DUSP7 is frequently over-expressed (Levy-Nissenbaum et al., 2003), but its physiological role is still unknown.
We show here that DUSP7 is essential for two crucial steps of oocyte meiosis: first, it is required for timely NEBD to facilitate the reinitiation of meiosis; second, it is essential for accurate chromosome segregation. We establish the mechanisms by which DUSP7 fulfills these functions and attribute a physiological role to DUSP7 during development.

RESULTS

\textbf{DUSP7 Depletion in Mouse Oocytes Delays and Prevents Meiotic Resumption}

We depleted DUSP7 by culturing small interfering RNA (siRNA)-injected oocytes within follicles in vitro (Figure S1A). This system is particularly well suited to deplete stable proteins in mouse oocytes, because it prevents protein synthesis from an early stage of oocyte growth onward (Pfender et al., 2015). In short, small immature oocytes that are surrounded by cumulus cells (follicles) were isolated from female mice around 11 days after birth. After microinjection with siRNAs against DUSP7 and subsequent culture for 10 days ex vivo, the fully grown and mature oocytes were stripped of the cumulus cells and microinjected a second time with mRNAs encoding fluorescent reporter proteins and rescue constructs. We then followed meiosis by high-resolution live cell microscopy.

First, we verified that our follicle culture system is suitable to study the regulation of cell-cycle events by comparing in vivo and in vitro grown oocytes. In both groups, the activity of the E3 ubiquitin ligase anaphase promoting complex/cyclosome (APC/C) was comparable (Figure S1B). In addition, the levels of E3 ubiquitin ligase anaphase promoting complex/cyclosome and in vitro grown oocytes. In both groups, the activity of the study the regulation of cell-cycle events by comparing in vivo constructs. We then followed meiosis by high-resolution live cell microscopy.

We depleted DUSP7 by culturing small interfering RNA (siRNA)-injected oocytes within follicles in vitro (Figure S1A). This system is particularly well suited to deplete stable proteins in mouse oocytes, because it prevents protein synthesis from an early stage of oocyte growth onward (Pfender et al., 2015). In short, small immature oocytes that are surrounded by cumulus cells (follicles) were isolated from female mice around 11 days after birth. After microinjection with siRNAs against DUSP7 and subsequent culture for 10 days ex vivo, the fully grown and mature oocytes were stripped of the cumulus cells and microinjected a second time with mRNAs encoding fluorescent reporter proteins and rescue constructs. We then followed meiosis by high-resolution live cell microscopy.

Next, we wanted to identify the targets of DUSP7 that drive the reinitiation of meiosis. To identify these substrates, we carried out an in vitro phospho-peptide screening assay. In this assay, synthetic 12–13 amino acid-long peptides with a phosphorylated serine, threonine, or tyrosine residue are incubated with a phosphatase. If the phosphatase dephosphorylates a peptide, the phosphate moiety is released and detected by a chelating solution that changes color upon phosphate binding. We purified active DUSP7 from bacteria and performed five independent in vitro dephosphorylation assays. We then focused on protein substrates that were found in at least three of the assays (Figure 2A). In this way, we identified Ephrin type-A receptors (Epha), phospholipase C gamma 1 (PLCγ1), and different conventional protein kinase C (cPKC) isoforms as DUSP7 substrates. RNAi-mediated targeting of these proteins did not result in a detectable phenotype (Figures S2A–S2C). However, previous studies suggested that activation of cPKC isoforms, for example, by Phorbol 12-myristate 13-acetate (PMA), a highly potent activator of cPKC isoforms, delays meiotic resumption in mouse oocytes (Downs et al., 2001; Lefèvre et al., 1992). We confirmed that PMA delays meiotic entry, mimicking the DUSP7 depletion phenotype (Figures 2B–2D). In addition, the specific upregulation of cPKC activity by overexpressing constitutively active PKCζ (Hennige et al., 2010) delayed NEBD to a similar extent as DUSP7 depletion (Figures S2D–S2F). Based on these results, we hypothesized that DUSP7 promotes NEBD by dephosphorylating and inactivating cPKC isoforms. Consequently, cPKC isoforms should remain active in the absence of DUSP7. If this hypothesis holds true, inactivation of cPKC isoforms in DUSP7-depleted oocytes should rescue the
depletion phenotype. We thus treated DUSP7-depleted oocytes with the cPKC inhibitor CGP-53353 (Traxler et al., 1997). Increasing concentrations of the inhibitor partially rescued the number of oocytes that underwent NEBD (Figure 2E). To confirm the specificity of this result, we co-depleted DUSP7 and PKCβ in follicles. In addition, the co-depletion of PKCβ rescued the efficiency of NEBD in DUSP7-depleted oocytes (Figures S2H–S2L). In both rescue conditions, the timing of NEBD was still slower than in controls (Figure 2F; Figure S2I), which is likely due to increased, but overall lower, levels of Cdk1/CycB activity, as we discuss in more detail later.

To demonstrate the physiological relevance of these findings, we tested in vivo whether cPKC activity is increased in DUSP7-depleted oocytes. As a measure of cPKC activity, we used an
antibody against auto-phosphorylated cPKC, because the degree of auto-phosphorylation represents an estimate of cPKC activity (Edwards et al., 1999). In particular, we used an antibody specific for phosphorylated threonine 641 of PCK(2) (pT641), which was also identified as DUSP7 target site by the in vitro phospho-peptide assay. We verified that this antibody is phospho-specific (Figure S2M). We found that the pT641 signal is increased in DUSP7-depleted compared to control-depleted oocytes (Figures 2G and 2H). Moreover, in DUSP7-depleted samples only, we observed a second band above the phospho-PCK(2) signal, which might represent a highly phosphorylated, active form of cPKC (Edwards et al., 1999). The total protein level of PCK(2) was unchanged, as judged by immunoblotting with a PKC(2) antibody (Figure 2G). Collectively, these results establish that DUSP7 dephosphorylates cPKC isoforms in mouse oocytes to facilitate NEBD.

Restoring Cdk1/CycB Activity in DUSP7-Depleted Oocytes Rescues DUSP7 Depletion

Our in vitro phospho-peptide assay identified cPKC isoforms as DUSP7 targets. However, the mechanism by which DUSP7 depletion causes a prolonged meiotic arrest remained unclear. A prerequisite for NEBD is Cdk1/CycB activation. We speculated that the activity of Cdk1/CycB might be too low to support timely NEBD in DUSP7-depleted oocytes. We tested this hypothesis with a radioactive Cdk1 kinase assay using histone H1 as a substrate (Figure 3A; Figure S3A). In particular, we quantified Cdk1/CycB activity in mouse oocytes 6 hr after release from prophase arrest, because the Cdk1/CycB1 activity peaked at this time point in both conditions (Figure S3A). We found that Cdk1/CycB activity is reduced by around 40% in DUSP7-depleted oocytes (Figure 3B). In addition, we performed the assay in the presence of the cPKC activator PMA and observed a similar decrease in Cdk1/CycB activity (Figure 3C). We speculate that in the absence of DUSP7, Cdk1/CycB activity drops below the critical level required to resume meiosis. Expression of GFP-tagged wild-type DUSP7 restores Cdk1/CycB activity to levels that are sufficient to promote NEBD, but the overall level of activity may still be lower than in controls. This might be why DUSP7 expression or cPKC inhibition or depletion in DUSP7-depleted oocytes rescues the number of oocytes that undergo NEBD but does not restore normal timing of NEBD.

If the DUSP7 depletion phenotype is due to decreased Cdk1/CycB activity, increasing Cdk1/CycB activity should rescue the DUSP7 depletion phenotype. To test this, we artificially increased Cdk1/CycB activity to three levels (Figure 3D). First, we increased the amount of wild-type Cdk1 protein (Cdk1wt). This should only result in a mild increase in Cdk1/CycB activity, because the kinases WEE1 and MYT1 still phosphorylate and thereby inhibit Cdk1wt (Han et al., 2005; Oh et al., 2010). Expression of Cdk1wt did not change the percentage of oocytes that underwent NEBD (Figure 3E) but caused a minor reduction in the timing of NEBD in DUSP7-depleted oocytes (Figure 3F). Next, we expressed an active Cdk1 mutant that cannot be phosphorylated by WEE1 and MYT1 (Cdk1 T14A, Y15F: Cdk1AF). This should result in a strong increase of Cdk1/CycB activity (Pomerening et al., 2008). Control and DUSP7-depleted oocytes expressing GFP-Cdk1AF underwent NEBD shortly after release from prophase arrest, when EGFP-expressing oocytes were still arrested (Figure 3G). We obtained the same result when we expressed CDC25B, the phosphatase that activates Cdk1/CycB in mouse oocytes (Figure S3B) (Lincoln et al., 2002; Okazaki et al., 1996; Pirino et al., 2009). Both conditions increase Cdk1/CycB activity to very high levels (Figure 3D) and lead to a premature rescue of the DUSP7 depletion phenotype. Finally, we increased Cdk1/CycB activity to intermediate levels by expressing Cyclin B1, the co-activator of Cdk1 (de Vantéry et al., 1997). Expression of mCherry-tagged Cyclin B1 (CycB1-mCherry) in DUSP7-depleted oocytes was able to fully rescue the DUSP7 depletion phenotype. The percentage of oocytes that underwent NEBD and the timing of NEBD were comparable to the results for control oocytes (Figures 3I and 3J). This suggests that decreased Cdk1/CycB activity is the cause for the delay of NEBD in DUSP7-depleted oocytes

The Spindle Assembly Checkpoint Is Compromised in DUSP7-Depleted Oocytes

DUSP7 depletion caused a second prominent phenotype, in addition to impairing meiotic resumption: the DUSP7-depleted oocytes that underwent NEBD often had misaligned chromosomes close to anaphase (Figures 1A, inset, and 1E). In addition,
the time from NEBD to anaphase was drastically shortened (Figure 1F). These defects were specifically due to the depletion of DUSP7, because expression of wild-type (DUSP7wt) or ERK1/2 binding deficient (DUSP7MBM) DUSP7 partially rescued both defects (Figures 1E and 1F). The rescue required the catalytic activity of DUSP7, because catalytically inactive DUSP7 (DUSP7CA) did not rescue the defects (Figures 1E and 1F).

Chromosome alignment at the metaphase plate is preceded by the attachment of kinetochores to spindle microtubules. The spindle assembly checkpoint (SAC) delays anaphase onset in response to unattached chromosomes by generating an inhibitory signal (Kouznetsova et al., 2014; Touati and Wassmann, 2016; Wassmann et al., 2003). The kinetochore localized kinases MPS1 and Aurora B participate in both steps to promote the correct attachment and alignment of chromosomes on the metaphase plate. Inhibition of MPS1 has been reported to lead to chromosome misalignment and premature anaphase onset in mouse oocytes (Hached et al., 2011; Nijenhuis et al., 2014). Given that the depletion of DUSP7 results in chromosome misalignment and premature progression into anaphase, we investigated whether SAC signaling could be impaired in the absence of DUSP7. To test this possibility, we overexpressed a core component of the SAC: MAD1 (SNAP-tagged MAD1, SNAP-MAD1). MAD1 localizes to the kinetochores of unattached chromosomes, where it catalyzes the generation of the inhibitory signal (Kulukian et al., 2009; Sironi et al., 2001). Overexpression of SAC components in mouse oocytes causes a metaphase arrest independently of misaligned chromosomes if the SAC is functional (Wassmann et al., 2003). As the SAC component, we decided to overexpress SNAP-MAD1, because it is also suitable to mark kinetochores of misaligned chromosomes by live cell microscopy (Chmátal et al., 2015). The expression of SNAP-MAD1 in control or DUSP7-depleted oocytes did not influence the number of oocytes undergoing NEBD or the timing of NEBD (Figures S4A and S4B). As expected, control oocytes expressing SNAP-MAD1 arrested in metaphase (Figure 4B). In contrast, DUSP7-depleted oocytes expressing SNAP-MAD1 did not arrest in metaphase but progressed into anaphase after a delay (Figure 4B; Figure S4C).

We then investigated the localization of SAC proteins. Control oocytes, as well as DUSP7-depleted oocytes, showed a positive SNAP-MAD1 signal at kinetochores shortly after NEBD (Figure 4A; Figure S4D). This confirms that the initial recruitment of SAC components to the kinetochore is not altered in the absence of DUSP7. Misaligned chromosomes in control oocytes are rare and can only be observed transiently upon formation of a metaphase plate in control oocytes (Chmátal et al., 2015). However, previous studies showed that they are positive for MAD1 (Chmátal et al., 2015; Zhang et al., 2005). Consistent with these studies, 81% (9 of 11 recordings) of misaligned chromosomes in 33 control oocytes were positive for MAD1 (Figure S4E). By contrast, only 26% (5 of 19 recordings) of misaligned chromosomes in 28 DUSP7-depleted oocytes were positive for MAD1 (Figure S4E). These data suggest that SAC proteins are not efficiently retained at misaligned kinetochores in the absence of DUSP7. If this holds true, inactivating the SAC by other means should show a similar phenotype. Inhibition of the kinase MPS1 is known to disturb SAC signaling in mouse oocytes, and the oocytes progress into anaphase in the presence of misaligned chromosomes (Hached et al., 2011). Depletion of DUSP7 or MPS1 resulted in a similar time from NEBD to anaphase (Figure 4C). Co-depletion of MPS1 in DUSP7-depleted oocytes reduced the time of anaphase further, possibly due to a more complete inactivation of the checkpoint (Figure 4C). Both MPS1 and DUSP7 depletion caused chromosome alignment defects. The percentage of oocytes with misaligned chromosomes was higher in MPS1-depleted oocytes than in DUSP7-depleted oocytes and did not increase further upon co-depletion (Figure 4D). The number of oocytes undergoing NEBD and the time from release to NEBD were not influenced by MPS1 depletion (Figures S4F and S4G). However, the overall phenotypes were similar, consistent with impaired SAC signaling in DUSP7-depleted oocytes.

Figure 3. Increased Cdk1/CycB Activity Rescues DUSP7 Depletion
(A) Follicles were depleted of DUSP7 or microinjected with scrambled (scr) RNAi. After culture, the oocytes were released and snap frozen at the indicated time points. The auto-radiogram shows the incorporation of radioactive phosphate into histone H1. Coomassie brilliant blue (CBB) staining shows histone H1.
(B) The intensity of the radioactive histone H1 signal at 6 hr after release was measured and normalized to histone H1 protein. The experiment was repeated three times.
(C) Mature oocytes were isolated and released in the presence of either DMSO or the cPKC activator PMA. Oocytes were snap frozen at the indicated time points. The auto-radiogram shows the incorporation of radioactive phosphate into histone H1. CBB staining shows histone H1.
(D) Scheme showing the different constructs to modulate Cdk1/CycB kinase activity and the expected relative increase of Cdk1/CycB activity. A + sign indicates a mild increase, ++ indicates a medium increase and +++ stands for Cdk1/CycB hyper-activation.
(E) Follicles were depleted as described in (A) but microinjected a second time after isolation with H2B-mCherry and GFP or GFP-Cdk1 wild-type (Cdk1WT). After mRNA expression, the oocytes were released and imaged on a confocal microscope. The percentage of oocytes that underwent NEBD was scored at the end of the imaging. The experiment was repeated three times.
(F) Follicles were treated as in (E), and the time from release from prophase arrest to NEBD was measured.
(G) Follicles were treated as in (F). GFP-tagged hyper-active Cdk1 (Cdk1HAT) was microinjected in a separate group of oocytes. This group underwent NEBD before imaging at the microscope was started, and analysis of the timing was therefore not possible. Instead, all oocytes that underwent NEBD within a time frame of 60 min after release (and within 30 min intervals afterward) were grouped together. The cumulative percentage of oocytes that underwent NEBD after the indicated time is shown.
(H) Oocytes from (E) that performed NEBD were analyzed for the presence of misaligned chromosomes in the frame before the onset of anaphase.
(I) Follicles were treated as in (E) but microinjected with H2B-mCherry and CycB1-mCherry. After mRNA expression, the oocytes were released and imaged on a confocal microscope. The percentage of oocytes that underwent NEBD was scored at the end of the imaging. The experiment was repeated five times.
(J) Oocytes from (I) were analyzed for the time from release from prophase arrest to NEBD.
(K) Oocytes from (I) that performed NEBD were analyzed for the presence of misaligned chromosomes in the frame before the onset of anaphase.
Figure 4. DUSP7 Participates in the SAC Response in Mouse Oocytes

(A) Follicles were microinjected with scrambled (scr) or Dusp7 RNAi and cultured. The somatic cells were removed and the oocytes were microinjected a second time with H2B-mCherry to mark the chromosomes and SNAP-tagged MAD1 (SNAP-MAD1). After mRNA expression, the SNAP tag was labeled with SNAP-Cell 505 Star. The oocytes were released and imaged on a confocal microscope. Representative images of these movies are shown. Misaligned chromosomes are highlighted with white arrowheads. Time is given in hours and minutes. Scale bar represents 10 μm. The inset shows a magnification of one misaligned chromosome per group. The channel for SNAP-MAD1 in the inset was adjusted in brightness to visualize MAD1 better. The full movies are available as Movies S4 and S5. The experiment was repeated four times.

(B) Oocytes from (A) and additional oocytes treated in the same way but microinjected with EGFP alone were analyzed for their progression into anaphase.

(C) Follicles were microinjected with scrambled (scr), Dusp7, and/or Mps1 siRNAs and cultured. The oocytes were microinjected with H2B-mCherry and EGFP-MAP4. After mRNA expression, the oocytes were released and imaged on a confocal microscope. Their time from release from prophase arrest to anaphase was analyzed. The experiment was repeated four times.

(D) The same oocytes as in (C) were analyzed for the presence of misaligned chromosomes in the last frame before anaphase.

(E) Representative images from oocytes used in Figure 3I to illustrate the earlier onset of CycB1-mCherry degradation in DUSP7-depleted oocytes.

(F) The CycB1-mCherry fluorescence was measured in oocytes from Figure 3I as described in Lane and Jones (2014). The average fluorescence intensity at NEBD was set to 100%.
A key target in SAC signaling is the APC/C. The APC/C is a multi-subunit ubiquitin ligase that regulates the onset of anaphase by targeting the anaphase inhibitor securin and other M-phase substrates for degradation (Barford, 2011; Primorac and Musacchio, 2013). The SAC delays anaphase by inhibiting the APC/C until the chromosomes are correctly attached to the spindle. If SAC signaling is compromised in DUSP7-depleted oocytes, the APC/C should be inefficiently inhibited. Therefore, the APC/C should trigger the degradation of its substrates earlier than under normal conditions. A well-known substrate of the APC/C is CycB1, the co-activator of Cdk1. We expressed CycB1-mCherry in control and DUSP7-depleted oocytes (Figures S4E and S4F). As predicted, the APC/C was activated earlier in DUSP7-depleted oocytes, as reflected by a significantly earlier onset of CycB1-mCherry degradation.

The decreased activity of the SAC could also be a consequence of the lower Cdk1/CycB activity that we observed in DUSP7-depleted oocytes (Figure 3A) (Clijsters et al., 2014; McCloy et al., 2014; Rattani et al., 2014). However, upregulation of Cdk1/CycB activity or inhibition of cPKC isoforms did not restore normal chromosome alignment in DUSP7-depleted oocytes (Figures S3H and S3K; Figures S2G and S2J). In addition, the time from NEBD to anaphase was not rescued when Cdk1/CycB activity was increased (Figures S2K, S3C, and S3D). These results suggest that the defects in SAC signaling are not solely due to a decrease in Cdk1/CycB in the absence of DUSP7.

**DISCUSSION**

The tight and reliable control of meiotic resumption is crucial for reproduction. Although defects in meiotic resumption have been implicated in infertility (Hsieh et al., 2007; Hunt and Hassold, 2008; Mitri et al., 2014; Richards et al., 2002), the molecular mechanisms that drive the reinitiation of meiosis are still incompletely understood. Our results provide mechanistic insights into how the prophase arrest in oocytes is abrogated.

We identified an unexpected signaling pathway that is essential for the release from prophase arrest. In this pathway, DUSP7 drives meiotic resumption by dephosphorylating and thereby inactivating cPKC isoforms, which is essential for timely NEBD. This pathway is likely to be crucial for meiosis, because a DUSP7 knockout mouse created by the International Mouse Phenotyping Consortium has been reported to be infertile. The DUSP7-dependent pathway could act in parallel to the canonical Cdk1/CycB activation mechanism that has been implicated in the release from prophase arrest. In the canonical pathway, cyclic AMP-activated protein kinase A (PKA) activates WEE1, which inhibits Cdk1/CycB and thereby prevents meiotic resumption (Han and Conti, 2006; Oh et al., 2010). After the LH surge, PKA and consequently WEE1 are inactivated and Cdk1/CycB becomes active. Having two parallel pathways that control meiotic resumption may allow oocytes to trigger the release from prophase arrest more robustly and to integrate more signals before oocytes are allowed to resume meiosis. This may reduce the risk of precocious meiotic resumption and increase the chance of conception.

DUSP7 promotes the release from prophase arrest by dephosphorylating and thereby inhibiting cPKC isoforms, a prerequisite for the timely activation of Cdk1/CycB. But what is the link between cPKC isoforms and Cdk1/CycB? Protein kinase C (PKC)α and PKCβs are known to regulate the Cdk inhibitors p21(CIP) and p27(Kip1) (Detjen et al., 2000; Frey et al., 1997). p21(CIP) controls the G2/M transition in mouse spermatogenesis (Kaushal and Bansal, 2007), and p27(Kip1) is required for prophase arrest in Drosophila oogenesis (Hong et al., 2003). It is thus conceivable that DUSP7 could regulate Cdk1/CycB activity by suppressing an inhibitor of Cdk1/CycB, such as p21 and p27, via cPKC isoforms. In addition, cPKC isoforms can influence the activity of the Cdk1/CycB-activating phosphatase CDC25. They can do so by regulating the activity of CDC25 directly (Yano et al., 2013) or by suppressing its expression (Yoshida et al., 2001). DUSP7 may thus be essential for the timely activation of CDC25.

In addition to a delay or a block in NEBD, we frequently observed misaligned chromosomes and premature progression into anaphase in oocytes depleted of DUSP7 (Figure 4). The SAC arrests mouse oocytes in metaphase until the chromosomes are attached to the meiotic spindle (Wassmann et al., 2003). However, evidence suggests that the SAC in mammalian oocytes is not as stringent as in mitotic cells and some misaligned chromosomes might remain undetected (Kitajima et al., 2011; Kolano et al., 2012; LeMaire-Adkins et al., 1997; Pfender et al., 2015; Zielsinska et al., 2015). Many proteins that participate in the SAC are regulated by phosphorylation (Funabiki and Wynne, 2013; London and Biggins, 2014). Consequently, protein phosphatases play a major role in regulating SAC signaling. For example, protein phosphatase 1 removes the phosphorylation on the kinetochore component KNL1 to facilitate stable microtubule attachments and SAC silencing (Liu et al., 2010). In contrast, BubR1 needs to remain phosphorylated to recruit PP2A-B56, which in turn helps to stabilize microtubule attachments (Suijkerbuijk et al., 2012). It is tempting to speculate that DUSP7 contributes to some of these processes. In our in vitro phospho-peptide screen, we did not detect SAC components as potential DUSP7 substrates, even though some were included in the assay (e.g., components of the RZZ complex). We also did not observe a localization of DUSP7 to kinetochores. Instead, DUSP7 remained cytoplasmic in all cell-cycle phases (Pfender et al., 2015). This could suggest that DUSP7 participates not directly in the SAC but indirectly, for example, by regulating other protein kinases or phosphatases.

Alternatively, a weakened SAC response in DUSP7-depleted oocytes could be a consequence of decreased Cdk1/CycB activity (Clijsters et al., 2014; McCloy et al., 2014; Rattani et al., 2014). It was shown that lowering Cdk1/CycB activity leads to an earlier onset of anaphase, even in the presence of misaligned chromosomes. Increasing Cdk1/CycB activity in bub1Δ/Δ mice (Rattani et al., 2014) restored the function of the SAC but required a non-degradable version of Cyclin B, artificially enhancing Cdk1/CycB activity. We were able to rescue the timing of NEBD in the absence of DUSP7 by increasing Cdk1/CycB activity using wild-type Cyclin B1 but could not restore a normal SAC response. While these results indicate that the decrease in Cdk1/CycB activity is not the primary cause of the impaired SAC response in the absence of DUSP7, a further increase in Cdk1/CycB activity might restore a normal SAC function.
DUSP7 is also expressed in human oocytes (Madissoon et al., 2014; Zhang et al., 2009), suggesting that its function may be preserved. An important factor that contributes to female infertility in humans is the inability of an oocyte to resume meiosis (Albertini et al., 2003; Hunt and Hassold, 2008; Hutt and Albertini, 2007). It is likely that the malfunction of proteins that are only essential in the oocyte, not in other parts of the body, contributes to this defect. DUSP7 is an excellent candidate for such a protein, because DUSP7 mutant mice are viable but infertile, suggesting that DUSP7 functions primarily in the germline. Analysis of DUSP7’s function in humans will be crucial for understanding whether this pathway for meiotic resumption is conserved and whether defects in DUSP7 could contribute to infertility.

**EXPERIMENTAL PROCEDURES**

**Preparation and Culture of Follicles, RNAi Sequences, and Drug Treatment of Oocytes**

All mice were maintained in a specific pathogen-free environment according to UK Home Office regulations. The work with mice was carried out at the Medical Research Council (MRC) Laboratory of Molecular Biology following the specifications of the UK Home Office. The project license was approved by the UK Home Office and evaluated by the MRC Cambridge Ethics Committee. The preparation of follicles and their culture is described in Pfender et al. (2015). RNAi depletion was performed for 10 days using a final concentration of 50 nM short interfering RNA oligonucleotide (oligo) in the oocyte. The RNAi oligo sequences for DUSP7, Epha4, and Mps1 are described in Pfender et al. (2015). PLCγ1 was depleted with an oligo mix targeting the sequences 5′-ACGGAGAAGACCTTCCAGGTCAA-3′, 5′-CAGGCGCTAGGGAAAGATCAA-3′, 5′-CAGGTCATGACTTTGTTCTA-3′, and 5′-CAGGAGAGAAGATCCTTATTA-3′. PKCβ isofoms 1 and 2 were depleted with an oligo mix targeting the sequences 5′-AAGGACCTGATTCTACATTA-3′, 5′-CCAGTCAATCATGAACACACA-3′, 5′-CTGTCGTCTTTGTGTACACACA-3′, and 5′-CCGGAGCAAACACAAGTTTAA-3′. All RNAi oligos were obtained from QiAGEN.

Oocytes treated with PMA or CGP-53353 (Sigma) were incubated with the drug for 1 hr before release. During release and imaging, the drug was still present.

**Confocal Microscopy and Image Preparation**

Confocal microscopy was performed as described in Pfender et al. (2015). All oocytes were recorded at least for 14 hr. For some movies, a Leica SP8 inverted confocal microscope equipped with a heated environmental chamber and a motorized stage was used. Imaging was performed with a Leica 40×/1.1 numerical aperture (NA) water immersion objective. All movies were analyzed and prepared using ImageJ (NIH) and Photoshop (Adobe). If the contrast or the brightness were adjusted, the procedure was applied to the whole image or image sequence. The intensity of bands from auto-radiograms or immunoblots was measured using ImageJ and corrected for background signal and loading control.

**mRNA Preparation and Injection**

mRNA was prepared with the mMessage mMachine Kit (Ambion), precipitated with LiCl, and taken up in 20 µL of RNase-free water. Quantitative microinjection was performed as described (Lafte et al., 2009). After injection of mRNAs into oocytes, expression was performed for at least 4 hr at 37°C.

**Immunoblotting**

All samples were mixed with NuPAGE sample loading buffer (Invitrogen) and heated at 95°C for 5 min. Antibodies used for western blotting were anti-FLAG M2 (mouse, Sigma), anti-ERK2 (rabbit, Santa Cruz), anti-α-tubulin (rat, Ab2 Serotec), anti-PI3Kα C-terminal (rabbit, Santa Cruz), anti-CDK1 (mouse, Santa Cruz), anti-CD25B (rabbit, Cell Signaling Technology), and anti-pT641 PKCα (rabbit, Cell Signaling Technology). All primary antibodies were used in 5% dry milk in PBS with 0.1% Tween 20 and incubated overnight at 4°C.


